Physico-chemical properties and consumer acceptance of instant cowpea (Vigna unguiculata) powder for complementary food

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Two cultivars of cowpea (white and brown testa coloured) were processed by pre-cooking for 10min. and dry-toasting (170°C, 10min) to render them ready-to-eat and to inactivate the anti-nutritional factors. The samples were dry-milled into powders. Ogi, traditional gruel from maize, was produced into powder. Five different blends of ogi and cowpea powder were prepared. Proximate chemical composition including Trypsin inhibition activity of the various blends was investigated. Also functional characteristics of the samples including bulk density, gelation capacity and water absorption capacity were investigated. Ogi supplemented with cowpea powder and 100% ogi were prepared into porridges which were presented for sensory evaluation among members of the university community. Each sample was evaluated for colour, taste, mouth-feel, flavour and overall acceptance. Moisture content values of the processed cowpeas ranged from 6.05 to 7.22% for boiled white cowpea and boiled brown cowpea respectively. Protein content ranged between 22.8% in the boiled white cowpea and 25.4% in the raw brown cowpea. There were 88.6-93.3% and 77.7-88.6% reduction in Trypsin inhibitor activity (TIA) of both white and brown cultivars of cowpea respectively. Sensory evaluation of ogi supplemented with cowpea revealed that supplementation at level above 30% is not acceptable among the panel. Brown cultivar of cowpea was preferred to the white cultivar.

Keywords: Cowpea flour, ogi flour, supplementation, antinutritional factor, consumer acceptance.

INTRODUCTION

In developing countries, especially African countries, malnutrition remains a major health problem in infants and pre-school children. The weaning period is the most critical in the life of infants and pre-school children with serious consequences for growth, resistance to diseases, intellectual development and survival if the child's nutritional needs are not met. Consequently, considerable efforts to improve health and nutritional status of growing children have focused on the production of nutritious low-cost complementary foods (Osundahunsi and Aworh, 2003). Unfortunately, in African countries, traditional infant foods are made from cereals, starchy roots and tubers that provide mainly carbohydrates with low quality proteins. These infant foods, exemplified by 'ogi', a fermented gruel or porridge made from maize, sorghum or millet, are the leading cause of protein-energy malnutrition in infants and pre-school children in Africa (Osundahunsi and Aworh, 2001). Cereals, in general, are low in protein and are limiting in some essential amino-acids, notably lysine and tryptophan. Supplementation of cereals with locally available legumes that are high in protein and lysine, although often limiting in sulphur-containing amino-acids, increases protein content of cereal-legume blends and their protein quality through mutual complementation of their individual amino-acids. Unfortunately, the presence of naturally occurring nutritional stress factors including protease inhibitors, haemagglutinins, tannins and phytoestrogens impairs the utilization of legumes (Osundahunsi and Aworh, 2003). Thus, there is the need to process legumes for good digestibility. Cowpea, commonly known as beans, is a major source of protein in Nigerian diet. Because of its importance in the diet of most developing countries, cowpea has been accorded priority research

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status by the Protein Advisory Group of FAO/UNO (Phillips, 1982). Cowpea generally undergoes one form of processing or the other to convert it into consumable forms and achieve the necessary digestibility and palatability. Considerable interests have evolved in expanding the use of cowpea in other forms such as production of ready-to-use (semi-instant) powder (McWatters, 1983; Ngoddy et al., 1986; Olapade et al., 2003; Olapade et al., 2004; Olapade et al., 2005). The aim of this study was to produce ready-to-eat (instant) cowpea powder for complementary food for infants.

MATERIALS AND METHODS

Procurement of materials

The two cultivars of cowpea seeds used for this study, IT97K-499-3S (white-coloured testa) and IT98K-131-2 (brown-coloured testa) were obtained from International Institute of Tropical Agriculture (IITA) in Ibadan. The seeds were cleaned, packaged in moisture proof polyethylene bags and stored under refrigeration condition (4°C) until used. White cultivars of maize and granulated sugar were obtained from Bodija market in Ibadan.

Preparation of instant cowpea powder

Each cultivar was divided into three batches each of 500 kg. The first batch was steeped in 1500 ml distilled water for 10 min and wet-dehulled. The cotyledons were then cooked in 1500 ml distilled water for 10 min and the cooking broth was discarded. The beans were dried in a cabinet drier at 60 °C for 8 h and then milled and sieved through 200 μm to obtain cooked cowpea powder. The second batch was conditioned by adding 10% distilled water, then allowed to equilibrate in a moisture proof polyethylene bag for 4 h and dried at 65 °C for 4 h, then toasted for 10 min at 120 °C with constant stirring in an oven, allowed to cool, then cracked and winnowed. The cotyledons were milled and sieved through 200 μm to obtain toasted cowpea powder. The third batch was steeped in 1500 ml distilled water for 10 min and wet-dehulled. The beans were dried in a cabinet drier at 60 °C for 8 h and then milled and sieved through 200 μm to obtain raw cowpea powder.

'Gji' cake was prepared as described by Akingbala et al., (1981). The cake was dried at 50 °C for 2 h then milled into powder using a hammer mill through 0.8 mm screen.

Chemical analyses

Proximate chemical composition of the powders including moisture, crude fibre, fat, ash and crude protein (N x6.25), was determined using AOAC (2000) methods. Carbohydrate content was determined by difference. Trypsin inhibition activity was determined according to the modified method of Kakade et al. (1974).

Functional properties of powders

Functional properties of powders including foaming capacity, water absorption capacity, gelation capacity and bulk density were investigated as previously described (Olapade et al., 2003). Swelling power was determined as described by Leach et al (1959).

Preparation of complementary foods

Different blends of ogi and cowpea powder at 80:20, 70:30, 60:40 and 50:50 (w/w) respectively were prepared and thoroughly blended using a warring blender. Each blend (20 g) was reconstituted with 10 ml water and stirred into smooth slurry followed by addition of boiling water (40ml) and stirring to obtain consistent paste, which was sweetened with 5 g granulated sugar. Ogi (100%) was used as control.

Sensory evaluation

Coded samples were presented to a twenty-five member panel, which are familiar with the ogi for sensory evaluation. The panellists rated the colour, taste, flavour, mouth feel and overall acceptance using a nine point hedonic scale, where 9 indicated 'like extremely' and 1 'dislike extremely'.

Statistical analysis

Data obtained were tested for differences using Analysis of Variance and means were separated by Duncan's multiple range tests using SPSS 11.0 package.

RESULTS AND DISCUSSION

Chemical composition

The proximate composition of the cowpea powders was essentially similar (Table 1) and within the range generally reported for cowpea (McWatters 1983, Ngoddy et al. 1986, Olapade et al. 2003). There were significant differences in chemical composition between the cultivars used in this study (p<0.05). Protein, fat and crude fibres were higher in the brown cultivars while the white cultivar
Table 1 Proximate chemical composition of cowpea powders

<table>
<thead>
<tr>
<th>Sample</th>
<th>Moisture %</th>
<th>Protein %</th>
<th>Fat %</th>
<th>Crude fibre %</th>
<th>Ash %</th>
<th>Carbohydrate %</th>
<th>TIA IU/g sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raw white cowpea</td>
<td>11.2b</td>
<td>23.4cd</td>
<td>1.64c</td>
<td>1.89b</td>
<td>3.20b</td>
<td>58.7e</td>
<td>10.6b</td>
</tr>
<tr>
<td>Raw Brown cowpea</td>
<td>12.3a</td>
<td>25.4a</td>
<td>2.00b</td>
<td>2.40a</td>
<td>2.13d</td>
<td>56.1f</td>
<td>18.4a</td>
</tr>
<tr>
<td>Boiled white cowpea</td>
<td>6.05e</td>
<td>22.8d</td>
<td>0.84e</td>
<td>1.76b</td>
<td>4.34a</td>
<td>64.2b</td>
<td>0.71e</td>
</tr>
<tr>
<td>Boiled Brown cowpea</td>
<td>7.22c</td>
<td>24.9ab</td>
<td>1.12d</td>
<td>2.10ab</td>
<td>3.45b</td>
<td>61.2d</td>
<td>2.10d</td>
</tr>
<tr>
<td>Toasted white cowpea</td>
<td>6.70cd</td>
<td>23.9bc</td>
<td>0.97de</td>
<td>1.90b</td>
<td>2.50c</td>
<td>64.0b</td>
<td>1.21e</td>
</tr>
<tr>
<td>Toasted Brown cowpea</td>
<td>6.55cd</td>
<td>25.1a</td>
<td>1.01d</td>
<td>2.11ab</td>
<td>2.14d</td>
<td>63.1c</td>
<td>4.10c</td>
</tr>
<tr>
<td>Og1</td>
<td>6.57cd</td>
<td>9.81e</td>
<td>3.53a</td>
<td>1.81b</td>
<td>2.01d</td>
<td>75.3a</td>
<td>ND</td>
</tr>
</tbody>
</table>

Means are of triplicate trials
Means with the same subscripts in a column are not different (p<0.05)
ND= Not determined

was higher in ash. Trypsin inhibition activity (TIA) was higher in brown cultivar (Table 1). There were significant reductions in TIA as a result of the treatments (p<0.05). Cooking with water resulted in higher reduction of TIA than dry toasting of cowpea. Heat treatment especially cooking was reported to reduce Trypsin inhibition activity by 96-100% in legumes (Satwadhar et al. 1981, Chung et al. 1981, Adewusi and Osuntogun 1991, Apati and Ologhobo, 1997). Akinwale (1989) also reported 82% reduction in the TIA of cooked cowpea dishes. Bressani (1985) attributed loss of toxicity of protease inhibitory activity to the effect of heating. The proximate composition of the blends was not determined but it is expected that increase in level of cowpea substitution would result in increased protein content of the blends. Previous workers have reported increased protein content with increased legume substitution of cereals in complementary food formulations (McWatters, 1982; Ossai and Malomo, 1988; Ashaye et al., 2000; Obatol et al., 2000; Osundahunsi and Aworh, 2001).

Foaming capacity

Foaming capacity ranged from 6.3% for boiled white cowpea powder to 11.7% for toasted brown cowpea powder (Table 2). Boiling in water resulted in lower foaming capacity than toasting of cowpeas. There was no significant difference (p<0.05) between the cultivars used but difference was observed for the treatments used.

Water and fat absorption capacities

Water absorption capacity ranged from 121% in the boiled white cowpea powder to 162% in the toasted white cowpea powder. The fat absorption capacity was highest in brown cowpea (118%) and toasted white cowpea powder (117%). Boiling in water resulted in reduction of both water and fat absorption capacities of the cowpeas. Heat denaturation of cowpea protein has been associated with high fat absorption capacity of cowpea (Henshaw and Lawal, 1993). The results obtained were essentially within the range of earlier values reported for some cowpea flours (Olapade et al. 2003; Henshaw and Lawal, 1993).

Gelation capacity

The least gelation concentrations of the powders ranged from 17% to 19% (Table 2). The values obtained were similar to the values earlier reported for some cowpea flours (Abbey, 1988; Olapade et al., 2003).

Bulk density

Bulk density values ranged from 0.646 to 0.807g/ml for raw white cowpea and boiled brown cowpea powders respectively. Boiling in water resulted in greater value of bulk density for both cultivars used compared to toasting of cowpea. This may be attributed to partial gelatinisation of cowpea starch during boiling.

Swelling power

Swelling power values ranged from 3.07 in the boiled brown cowpea to 4.70 in the toasted brown cowpea powders. Toasting of cowpea produced greater swelling.
Table 2: Functional properties of cowpea powders

<table>
<thead>
<tr>
<th>Sample</th>
<th>Foam Capacity %</th>
<th>Water Absorption Capacity</th>
<th>Swelling Power %</th>
<th>Gelation Capacity %</th>
<th>Bulk Density g/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raw white cowpea</td>
<td>ND</td>
<td>131d</td>
<td>ND</td>
<td>ND</td>
<td>0.646c</td>
</tr>
<tr>
<td>Raw brown cowpea</td>
<td>ND</td>
<td>152b</td>
<td>ND</td>
<td>ND</td>
<td>0.680b</td>
</tr>
<tr>
<td>Boiled white cowpea</td>
<td>6.0</td>
<td>121e</td>
<td>3.33b</td>
<td>17b</td>
<td>0.790a</td>
</tr>
<tr>
<td>Boiled brown cowpea</td>
<td>7.0</td>
<td>123e</td>
<td>3.07b</td>
<td>18ab</td>
<td>0.807a</td>
</tr>
<tr>
<td>Toasted white cowpea</td>
<td>10.2</td>
<td>162a</td>
<td>3.67a</td>
<td>19a</td>
<td>0.650c</td>
</tr>
<tr>
<td>Toasted brown cowpea</td>
<td>12.0</td>
<td>147c</td>
<td>4.70a</td>
<td>17b</td>
<td>0.700b</td>
</tr>
</tbody>
</table>

Means are of triplicate trials
Means with the same subscripts in a column are not different (p<0.05)
ND – Not determined

Table 3: Sensory evaluation scores of complementary foods

<table>
<thead>
<tr>
<th>Percentage supplementation of instant cowpea</th>
<th>Colour</th>
<th>Taste</th>
<th>Flavour</th>
<th>Mouth-feel</th>
<th>Overall acceptance</th>
</tr>
</thead>
<tbody>
<tr>
<td>20% boiled white cowpea</td>
<td>7.7ab</td>
<td>8.2a</td>
<td>7.4bc</td>
<td>8.2a</td>
<td>7.9a</td>
</tr>
<tr>
<td>20% boiled brown cowpea</td>
<td>8.0a</td>
<td>8.1a</td>
<td>7.7ab</td>
<td>7.9a</td>
<td>7.9a</td>
</tr>
<tr>
<td>30% boiled white cowpea</td>
<td>7.3ab</td>
<td>7.9a</td>
<td>6.4d</td>
<td>8.0a</td>
<td>7.3b</td>
</tr>
<tr>
<td>30% boiled brown cowpea</td>
<td>6.7b</td>
<td>7.2a</td>
<td>6.2d</td>
<td>6.4c</td>
<td>6.1c</td>
</tr>
<tr>
<td>40% boiled white cowpea</td>
<td>5.3c</td>
<td>4.5b</td>
<td>5.4e</td>
<td>4.9de</td>
<td>5.1e</td>
</tr>
<tr>
<td>40% boiled brown cowpea</td>
<td>5.0c</td>
<td>5.4b</td>
<td>6.0d</td>
<td>5.2de</td>
<td>4.9e</td>
</tr>
<tr>
<td>20% toasted white cowpea</td>
<td>7.7ab</td>
<td>7.8a</td>
<td>8.0a</td>
<td>8.0a</td>
<td>8.1a</td>
</tr>
<tr>
<td>20% toasted brown cowpea</td>
<td>7.9a</td>
<td>8.1a</td>
<td>8.1a</td>
<td>7.9a</td>
<td>8.1a</td>
</tr>
<tr>
<td>30% toasted white cowpea</td>
<td>7.0ab</td>
<td>7.1a</td>
<td>7.1c</td>
<td>7.9a</td>
<td>7.1b</td>
</tr>
<tr>
<td>30% toasted brown cowpea</td>
<td>8.0a</td>
<td>7.4a</td>
<td>8.1a</td>
<td>8.2a</td>
<td>7.9a</td>
</tr>
<tr>
<td>40% toasted white cowpea</td>
<td>6.7b</td>
<td>5.4b</td>
<td>5.0e</td>
<td>5.4d</td>
<td>5.1e</td>
</tr>
<tr>
<td>40% toasted brown cowpea</td>
<td>6.7b</td>
<td>4.8b</td>
<td>5.4e</td>
<td>4.9e</td>
<td>5.4d</td>
</tr>
<tr>
<td>Ogi (control)</td>
<td>7.1ab</td>
<td>7.7a</td>
<td>7.2bc</td>
<td>7.1b</td>
<td>7.3b</td>
</tr>
</tbody>
</table>

Means with the same subscripts in a column are not different (P<0.05)

Power than boiling of cowpea. This may be attributed to partial gelatinisation of cowpea starch and denaturation of cowpea protein during boiling.

Sensory evaluation

Complementary foods containing 20% cowpea powder were comparable with ogi, the traditional weaning food, in sensory qualities (Table 3). Complementary foods containing 30% cowpea powder made from boiled beans received poorer scores for flavour (6.4 for white cowpea and 6.2 for brown cowpea) relative to ogi (7.2) and those made from toasted cowpea (7.1 for white cowpea and 8.1 for brown cowpea). Products containing 40% cowpea received significantly poorer scores for taste, flavour,
mouth-feel and overall-acceptability (Table 3).

CONCLUSION

Ogi can be enriched with toasted cowpea powder at a level of up to 30% without adverse effects on sensory qualities.

ACKNOWLEDGMENT

The authors wish to express their profound gratitude to Mr. Remi Adeleke of IITA Ibadan for the supply of cultivars of cowpeas used for this work. Also invaluable suggestion of Mr. Amaefula of IITA is highly appreciated.

REFERENCES